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INTRODUCTION

Proteasome inhibitors (PI) are currently at the backbone of treatment regimens of multiple myeloma (MM) therapy (1). Among them, bortezomib (BTZ) is the first-in class boronate-based PI approved for treatment of MM (2). Despite the improved outcomes brought by BTZ-based regimens, patients with MM relapse after BTZ treatment or become BTZ-refractory (2,3), a condition with a very poor prognosis (4). Thus, the development of the BTZ-resistance under continuing selective pressure of BTZ-containing treatments is an important clinical problem. Recent break-through studies unraveled the complexity of molecular mechanisms underlying the acquisition of BTZ-resistance (5,6). Thorough analysis of BTZ-resistant MM cells have revealed complex concerted changes in cell metabolism and protein turnover machinery, associated with myeloma cells becoming independent of proteasome machinery (5). Indeed, resistance to proteasome inhibitors in MM was found to be associated with an undifferentiated phenotype of plasmablasts and even earlier plasma cell progenitors (7), B lymphoid compartments implicated in extramedullary disease (8) and myeloma progeny (9). In fact, proteasome-independent protein turnover machinery relies on elevated activity of proteinases and peptidases (10). Noteworthy, elevated levels of peptidases have been noted in myeloma progression (see EP897), and can be targeted by a lipophilic peptide-conjugate melphalan flufenamide (hereafter referred to as melflufen) (11). Melflufen is a lipophilic peptide-drug conjugate of an alkylating moiety of melphalan (MPH) and a peptidase substrate. Due to its lipophilicity melflufen is easily transported through lipid membranes. Following transport, melflufen can be cleaved in peptidase-enriched compartments (endoplasmic reticulum, lysosomes, autophagosomes, etc.). This results in a dramatic decrease of lipophilicity and entrapment of alkylating moieties leading to enormous concentration of an alkylating agent within the peptidase-rich compartment. Melflufen has shown great potency in treatment-naïve and melphalan-resistant myeloma models (12, 13), and its efficacy in patients with RRMM has been documented (14). In this study, melflufen is under investigation in a BTZ-resistant MM setting.

OBJECTIVES

1. Evaluate efficacy of melflufen in BTZ-adapted/resistant (aBTZ) MM cell lines
2. Investigate the effect of melflufen on MM progenitor cells by clonal outgrowth
3. Analyze expression of peptidase-encoding genes in aBTZ MM cells
4. Assess the prognostic value of higher aminopeptidase expression in patients with MM

RESULTS

Figure 1. Cytotoxicity of melflufen in BTZ-adapted MM cells.

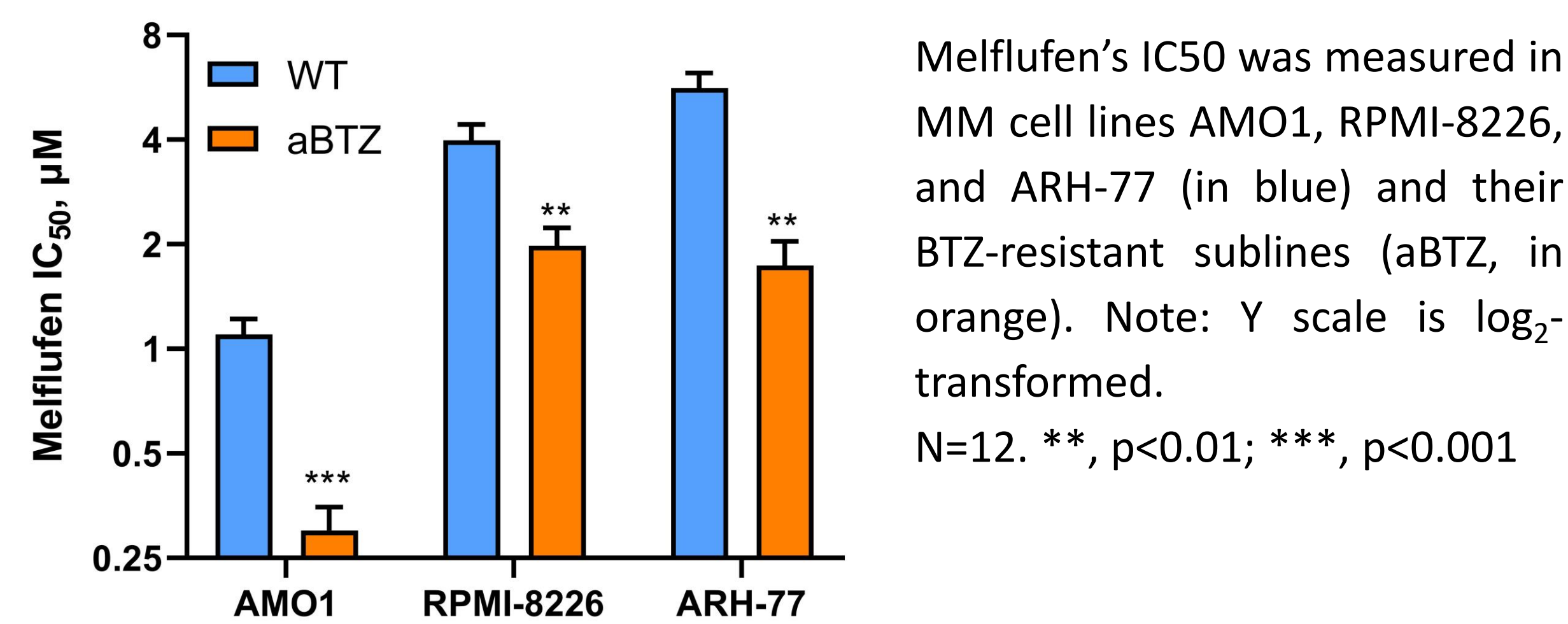


Figure 2. Apoptosis.

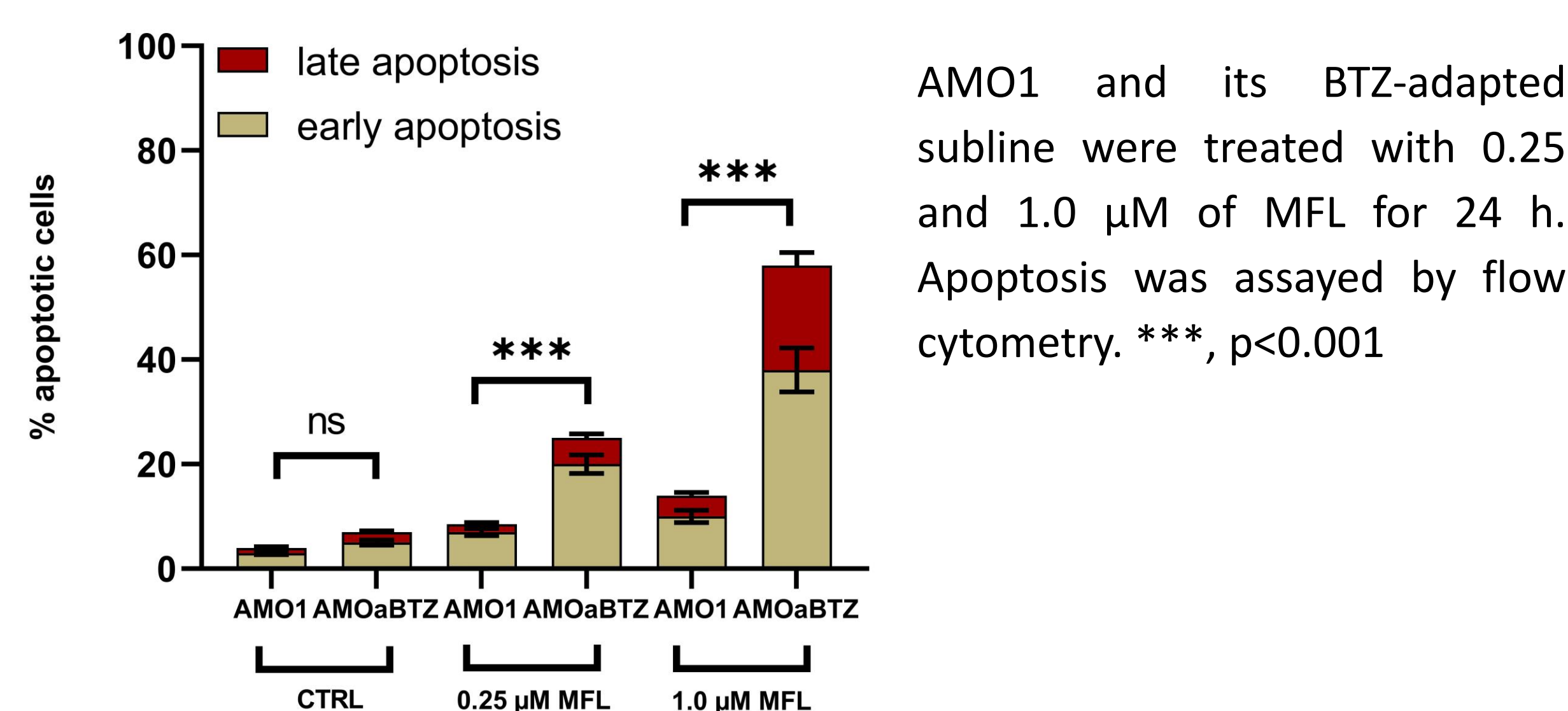


Figure 3. Expression of aminopeptidase genes *RNPEP* and *LAP3*.

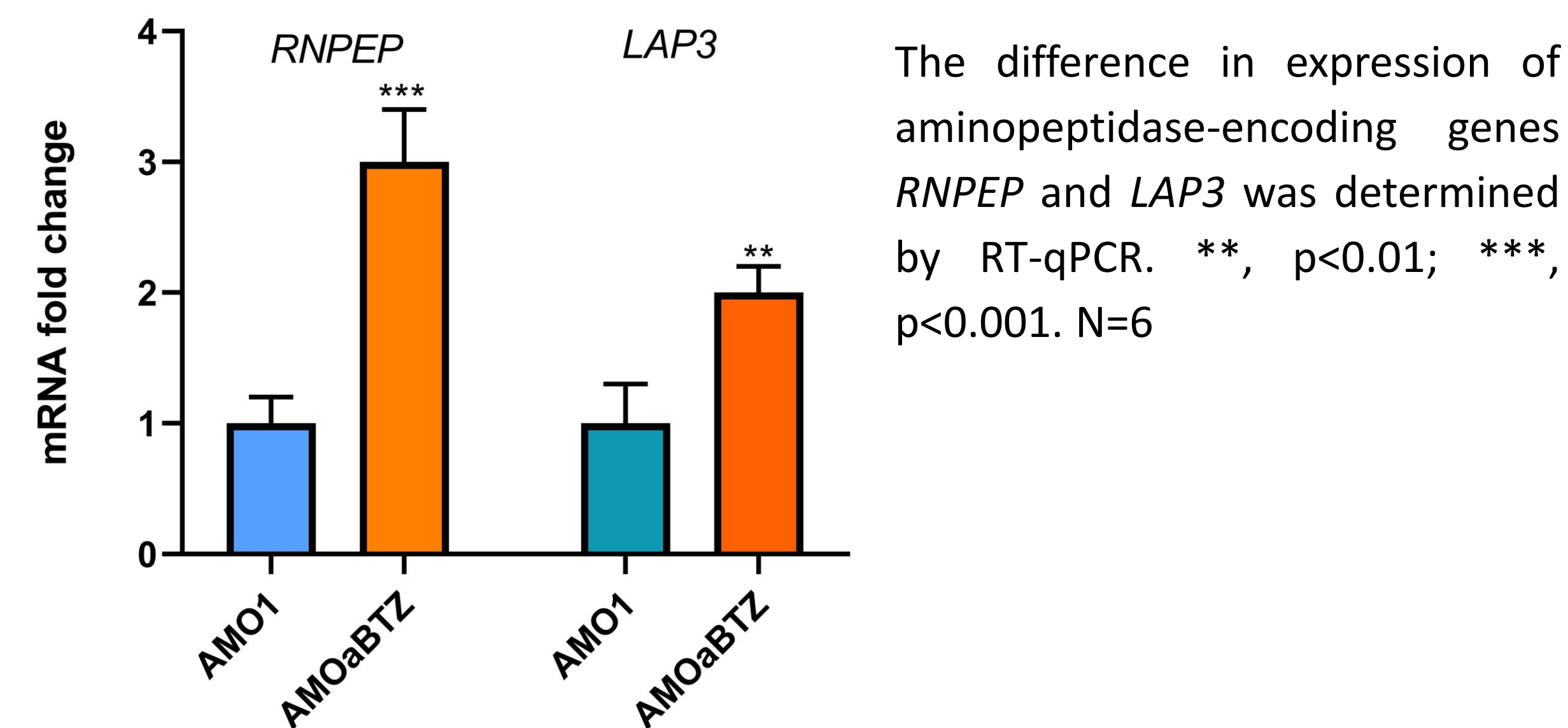


Figure 4. Effect of peptidase inhibitor bestatin (BST) on melflufen's cytotoxicity.

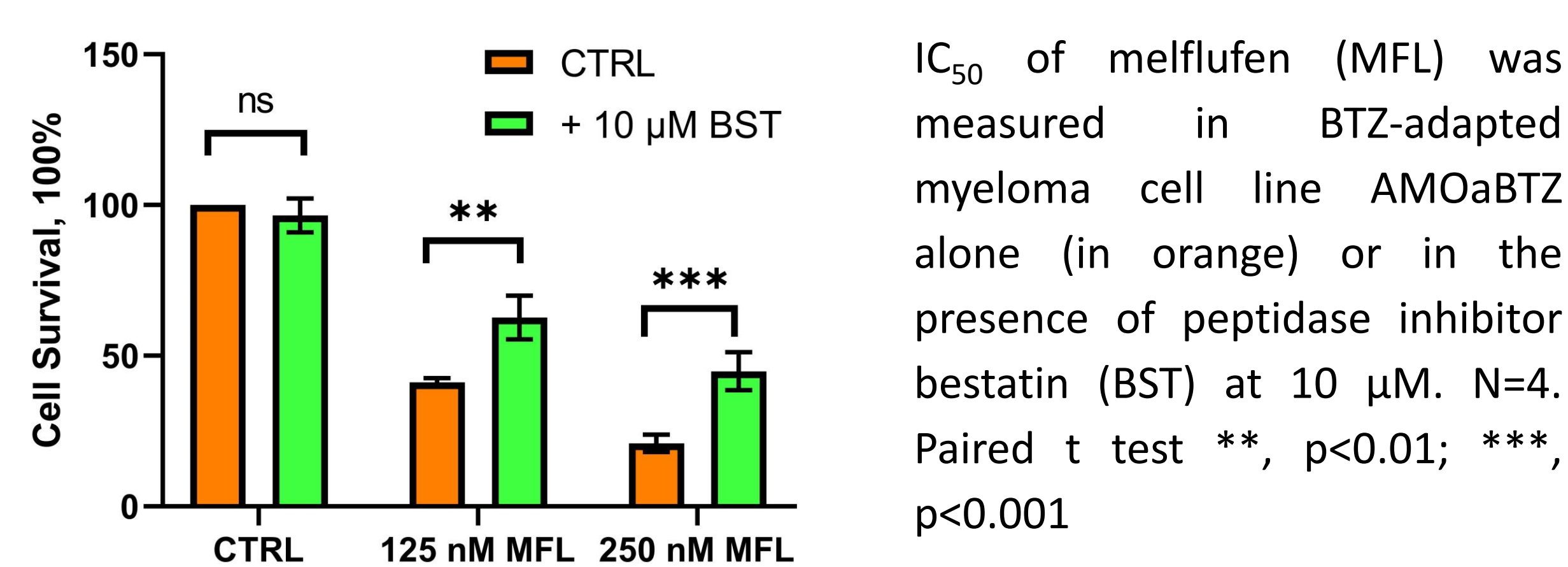


Figure 5. Aminopeptidase B expression in a murine myeloma model.

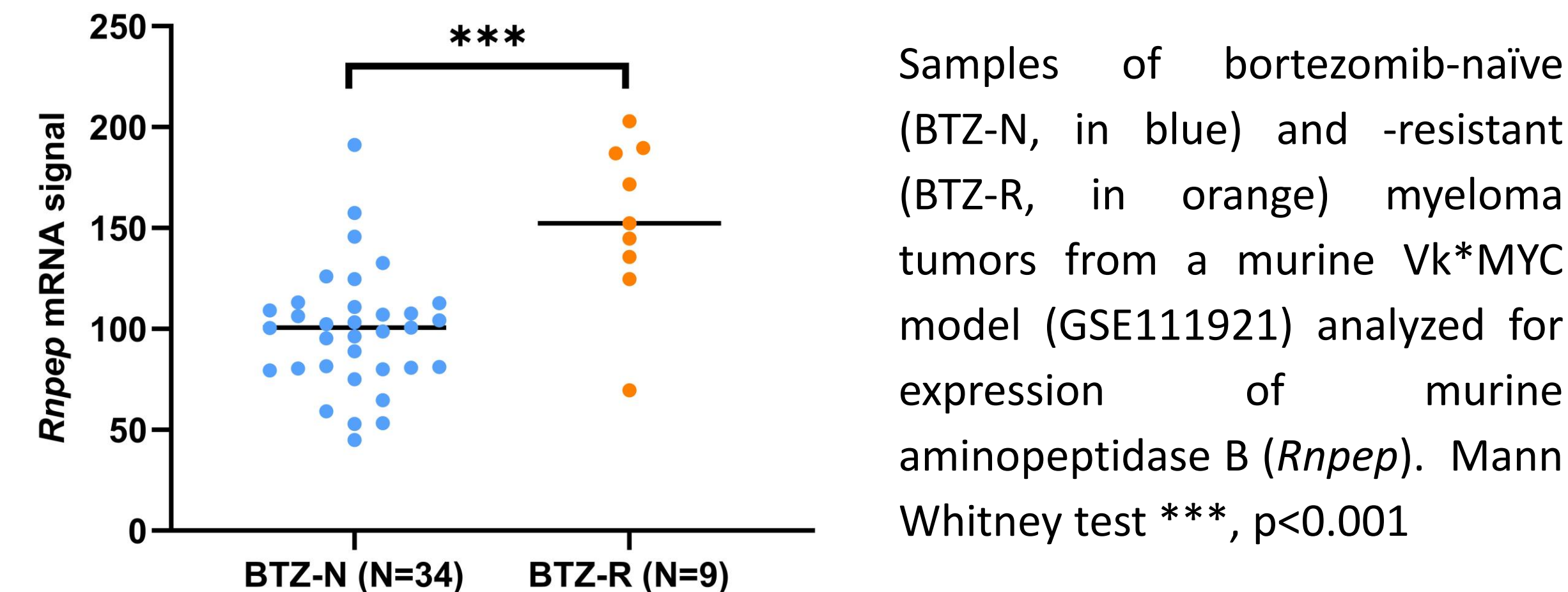


Figure 6. Progression-free survival (PFS) in MM patients treated by total therapy 3 (TT3).

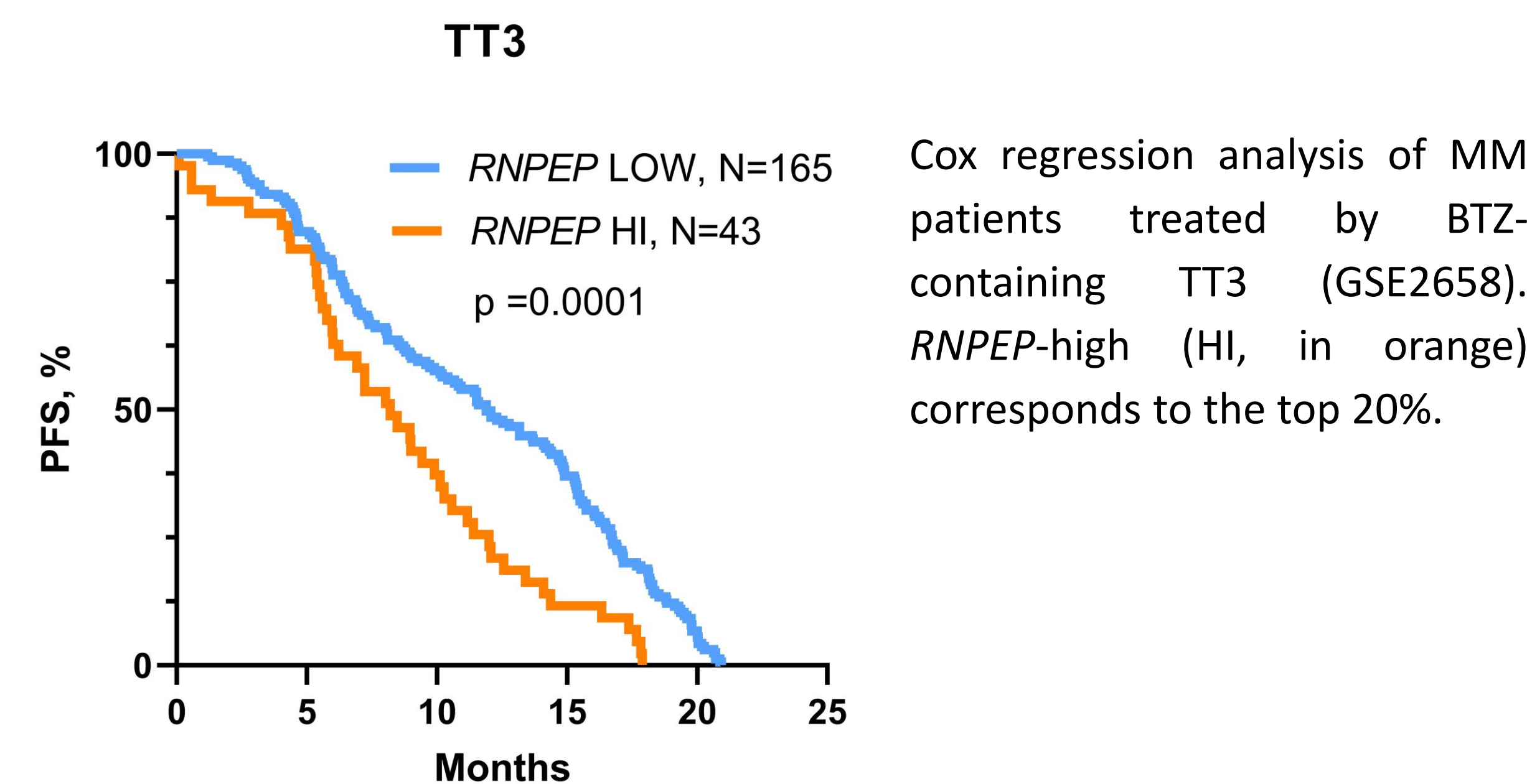


Figure 7. Clonal outgrowth of RRMM patient bone marrow samples.

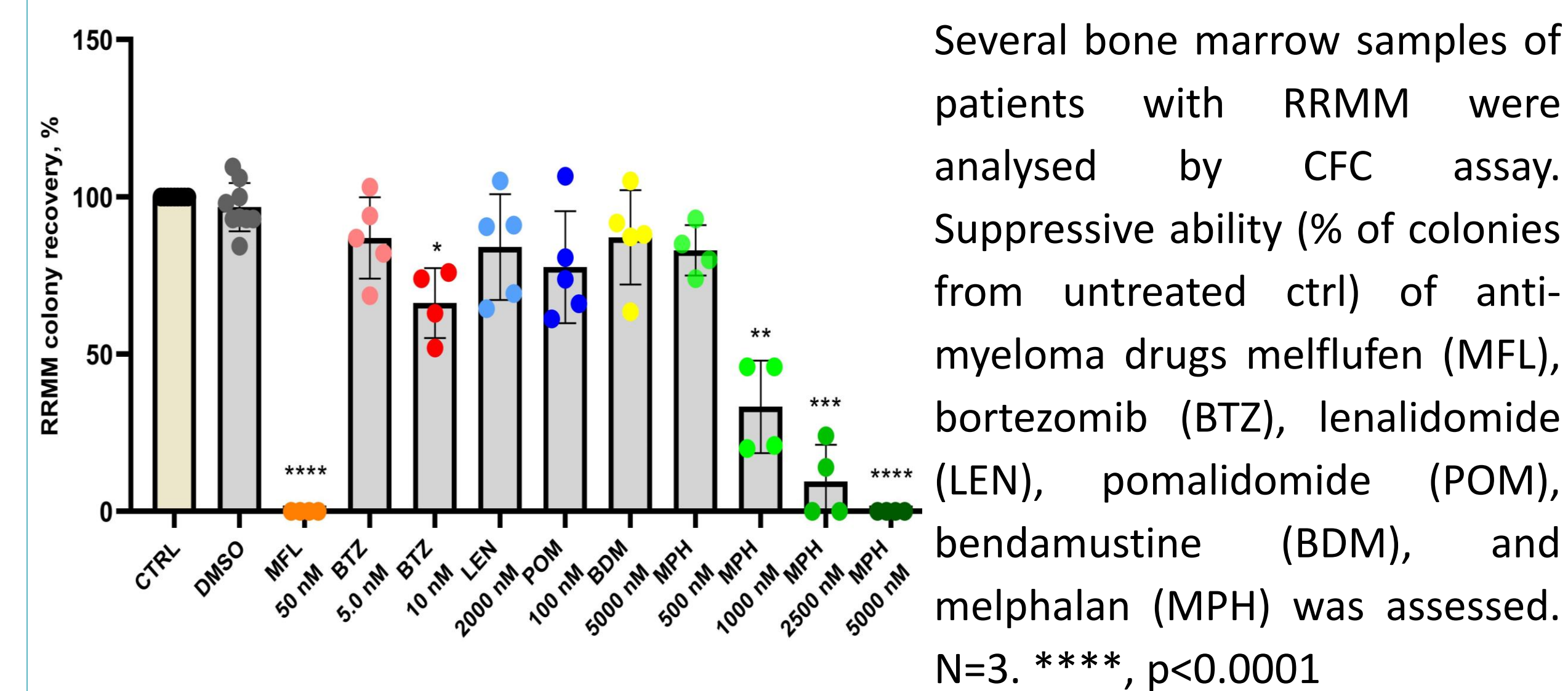
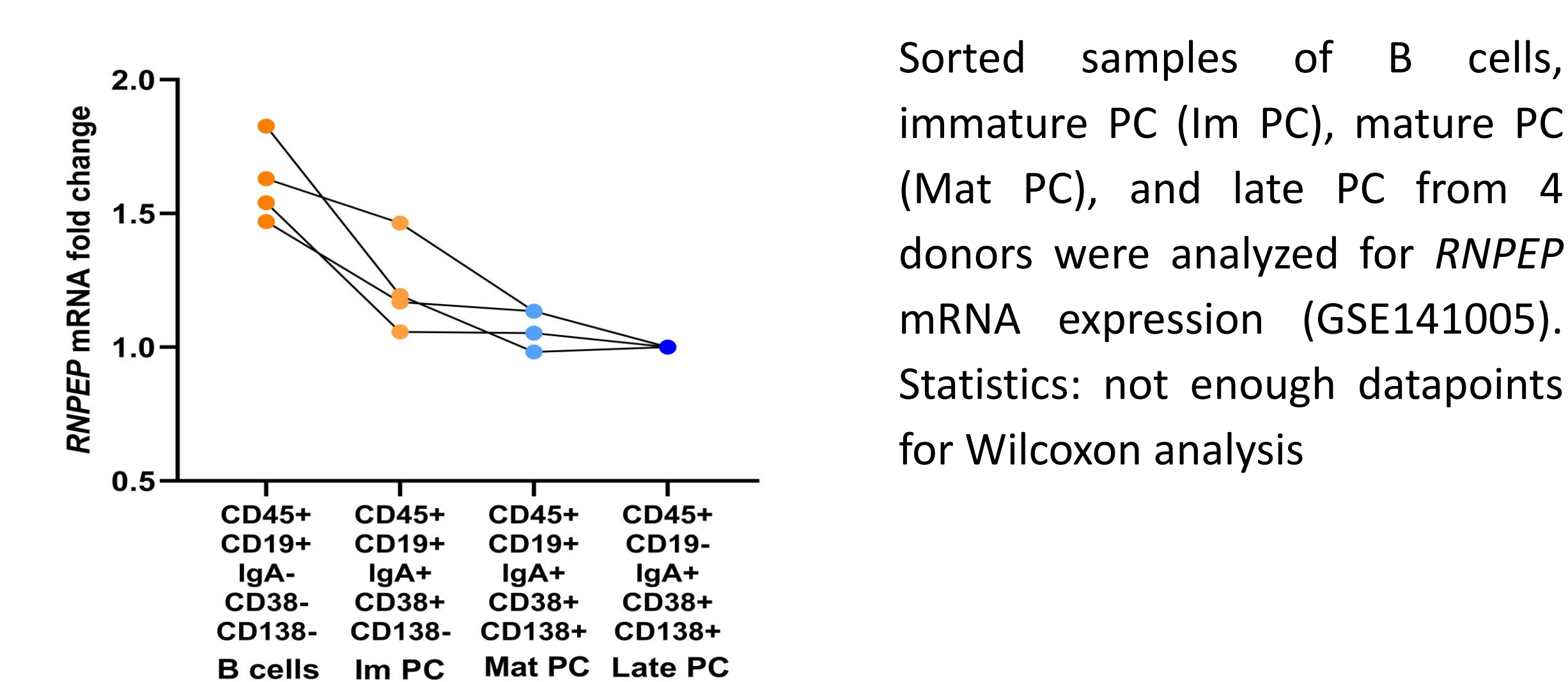
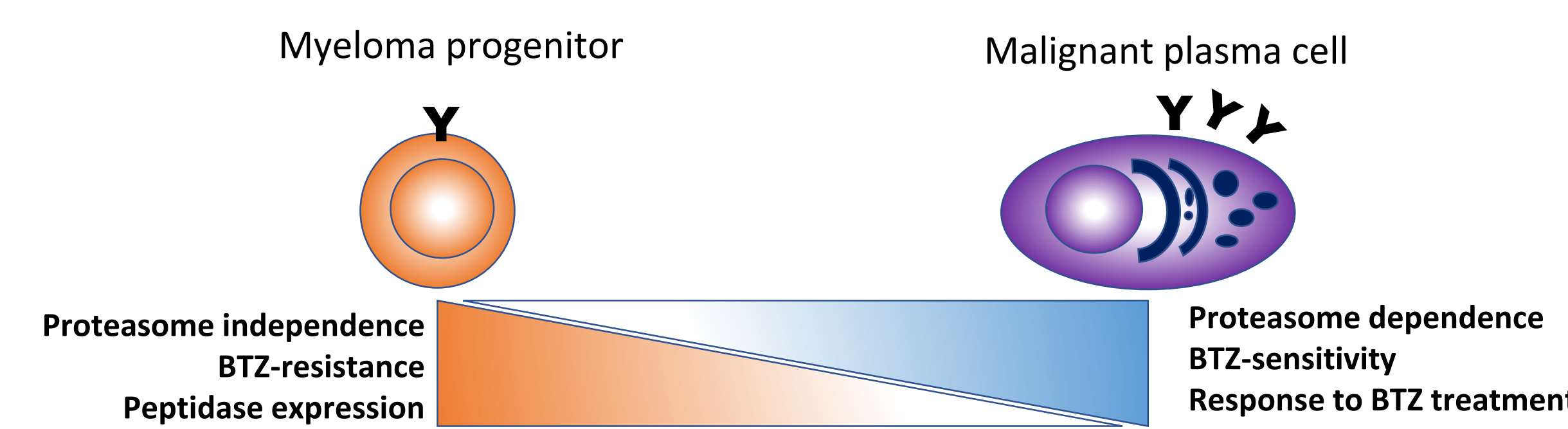


Figure 8. Aminopeptidase B expression in plasma cell (PC) differentiation.



CONCLUSIONS



Resistance of multiple myeloma to proteasome inhibitors is a multifaceted complex process involving metabolic adaptation, loss of proteasome dependence, and increased proteasome-independent protein turnover (5,6). Independence of proteasome activity implies increased peptidase activity involved in processing of unfolded proteins (10). In this study we have demonstrated that BTZ-resistant myeloma cells express higher levels of aminopeptidases *LAP3* and *RNPEP* in human and murine models. Higher peptidase expression confirmed higher sensitivity of BTZ-resistant cells to a novel peptidase-enhanced peptide-drug conjugate melflufen as shown by cytotoxicity assays in the presence of a peptidase inhibitor bestatin. As previously shown by gene set enrichment analysis (GSEA), resistance to proteasome inhibitors also coincides with undifferentiated plasma cell phenotype (7). Indeed, we were able to demonstrate that melflufen successfully suppresses clonal outgrowth of RRMM patients bone marrows. It implies potency of melflufen towards myeloma-initiating progenitor cells, a seed of disease relapse (9). This phenomenon is supported by an outstanding long overall survival of RRMM patients treated with melflufen in O-12-M1 study (14). Further investigation of melflufen's activity against myeloma progenitors and BTZ-resistant myeloma cells is currently ongoing.

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DISCLOSURES

KB, FL (Employment, Oncopeptides AB); FL (Equity, Oncopeptides AB); LB, CD (Research grant, Oncopeptides AB)